# Comparing the performance of diploid and triploid blue mussel (Mytilus edulis) larvae reared at constant and fluctuating temperatures



Author: Violet Chilvers Supervised by: Dr. Ramón Filgueira & Dr. Eric Ignatz

# INTRODUCTION

As anthropogenic climate change alters our ocean environment, a need to enhance hatchery-produced mussel seed is critical to sustain the aquaculture industry<sup>1,2</sup> and ensure sustainable food production<sup>3</sup>



- · Ectotherms exhibit natural intraspecific variation across geographic regions<sup>7,8</sup> due to either local adaptation<sup>9,10</sup> or developmental phenotypic plasticity<sup>9</sup> Carryover effect - early life thermal stress may enhance thermal tolerance later in
- Adult mussels from Sober Island Pond, NS, demonstrate > thermal tolerance compared to those from St. Peters, PE1

## **OBJECTIVES**

- To assess the performance (growth and survival) of blue mussel larvae across three factors: 1) ploidy - diploid vs. triploid 2) temperature regime - constant vs. fluctuating
  - 3) source NS vs. PE

# **HYPOTHESES**

Ploidy: triploids survival I particularly under thermal stress<sup>6</sup>, but achieve f shell length by the end of the experiment<sup>4</sup>

Temperature: larvae under constant temperature regimes will have \$ survival and growth than fluctuating regimes<sup>10</sup>

Source: NS larvae will have survival and mean length > PE larvae<sup>11,12</sup>

#### METHODS



 Induced spawning of NS and PE mussels by heat shock method<sup>13</sup> Triploid groups subject to 8000 PSI for 6 min, 18 min post-fertilization<sup>14</sup>



Figure 1. Factorial design of source (NS vs. PE), ploidy (diploid vs. triploid), and temperature regime (constant 18 °C vs. fluctuating 16-20 °C) with half-sibling families included in triplicate per treatment

Larval Husbandr

- o Fed daily after D-stage metamorphosis using a target algae cell concentration
- o Flask water changed, growth and survival of larvae assessed every other day







 Flow cytometry conducted at 8 and 20/22 days post-fertilization to determine percentage of triploids

Statistics Analysis Bayesian models



Figure 2. Effect of ploidy, temperature regime, and source on the survival of Mytilus edulis larvae from 2 to 22 days postfertilization. Line graphs show the survival of diploid and triploid larvae from Prince Edward Island at a constant temperature of 18 °C (A) and a fluctuating temperature of 16-20 °C (B), as well as for larvae from Nova Scotia under the same constant (C) and fluctuating (D) temperature conditions. The Bayesian model on day 22 suggests a lower survival in triploids with moderate certainty (UI: 72,71%), a negligible effect of temperature regime on survival (UI: 52,35%), and lower survival of Prince Edward Island mussels, with high certainty (UI: 96.38%).



Figure 3. Effect of ploidy, temperature regime, and source on the mean length of Mytilus edulis larvae from 2 to 22 days post fertilization. Line graphs show the mean length of diploid and triploid larvae from Prince Edward Island at a constant temperature of 18 °C (A) and a fluctuating temperature of 16-20 °C (B), as well as for larvae from Nova Scotia under the same constant (C) and fluctuating (D) temperature conditions. The Bayesian model of surviving families on day 22 indicates a lower mean length in triploids with a moderate certainty (UI: 69.19%), a lower mean length under fluctuating temperature with high certainty (UI: 95.96%), and a high certainty that PE larvae have a higher mean length (UI: 97.97%).



Figure 4. Percentage of triploid Mytilus edulis at 8 and 20/22 days post-fertilization across treatment groups with constant and fluctuating temperature regimes on larvae from Prince Edward Island and Nova Scotia sources. The Bavesian model indicates a higher triploid percentage in Prince Edward Island larvae with moderate certainty (UI: 80.46%), a reduction in triploidy occurrence under fluctuating temperature conditions with high certainty (UI: 98.55%), and a reduction in triploidy percentage from day 8 to day 20/22 with high certainty (UI: 96.07%).

# DISCUSSION

Ploidy: triploids survival I particularly under thermal stress, but 1 shell length by the end of the experiment

- Fail to reject Triploidy survival ↓ diploid survival<sup>6</sup>
- Reject Triploid length < diploid length<sup>5,15</sup>
  - Possibly due to lack of sexual maturation<sup>5,16</sup>

Temperature: larvae under constant temperature regimes will have 🕇 survival and growth than fluctuating regimes<sup>10</sup>

- Reject Survival is similar independently of temperature regime<sup>1</sup> However triploid individuals are decimated, reducing % triploids · Possibly due to differences in protein expression
- Fail to reject Growth is ↓ under the fluctuating temperature regime

Source: NS larvae will have survival and mean length > PE larvae11,12

- Fail to reject NS survival > PE
- Reject PE length and triploidy rates > NS.
  - Due to potential physiological differences<sup>7</sup>

# CONCLUSION

This study is an initial step to inform hatchery techniques that improve mussel performance under variable thermal conditions, with the goal of developing a resilient hatcherv-produced seed

#### Key conclusions

- o Triploids should maintain a constant hatchery standard temperature (~18°C)
- o Triploids do not obtain higher growth rates during larval stages
- Diploids are unaffected by thermal fluctuations compared to triploids

#### Future studies

- Study potential carryover effects of fluctuating treatments among diploid groups<sup>12</sup> Determine when triploid size surpasses diploids
- Further compare larval thermal resistance at iuvenile stages

### ACKNOWLEDGEMENTS

I would like to thank my supervisors, Dr. Ramón Filgueira & Dr. Eric Ignatz, my lab members Flavie Perron and Shelby Clarke, the Aquatron team and volunteers, Sobey Faye and Atlantic Aqua Farms for the funding and my friends and family: all of whom my research would not have been possible without

# REFERENCES

- 1. Fisheries and Oceans Canada. 2022. Accessed November 14th, 2024. https://www.dfo-
- Transformation of the second operational for the second operation of the second operation of the second operation of the second operation opera

22-eng.html.

- Willer DF. Aldridge DC. 2020. Nat Food. 1(7):384–388. doi:10.1038/s43016-020-0116-8.
- 4. Osterheld K. Davidson J. Comeau LA, Hori T. Belzile C. Tremblav R. 2023a, J Appl Aguac, 35(2):473-488. doi:10.1080/10454438.2021.1985032.
- . Brake J, Davidson J, Davis J. 2004. 236(1-4):179-191. doi:10.1016/j.aquaculture.2003.09.016.
- Li Y, Jiang K, Li Q. 2022. Aquaculture. 555:738219. doi:10.1016/j.aquaculture.2022.738219.
  Casas SM, Filgueira R, Lavaud R, Comeau LA, La Peyre MK, La Peyre JF. 2018. J Exp Mar Biol Ecol. 506:82–90.
- doi:10.1016/i.iembe.2018.06.001.
- Angilletta MJ. 2004. Integr Comp Biol. 44(6):498–509. doi:10.1093/icb/44.6.49
  Anestis A, Lazou A, Pörtner HO, Michaelidis B. 2007. Am J Physiol-Regul Integr Comp Physiol. 293(2):911–921.
- Donelan SC, Breitburg D, Ogburn MB. 2021. Ecol Appl. 31(4):e02315. doi:10.102/eap.2315
- Clarke SB, Ignatz EH, Hori TS, Comeau LA, Filgueria R. Halifax (NS): Dalhousie University.
  Sinclair BJ, Marshall KE, Sewell MA, Levesque DL, Willett CS, Slotsbo S, Dong Y, Harley CDG, Marshall DJ, Helmuth
- BS. et al. 2016. Vasseur D. editor. Ecol Lett. 19(11):1372-1385. doi:10.1111/ele.12686.
- Utting SD, Spencer BE. 1991. Lab Leaft MAFF Fish Res. Lowestoft (68):31pp. https://www.cefas.co.uk/publications/lableaflets/lableaflet68.pdf.
- Perron F, Ignatz EH, Chilvers V, Durier G, Benfey TJ, Hori TS, Filgueria R. 2024. In Preparation.
  Talmage SC, Gobler CJ. 2011. PLoS ONE. 6(10):2941-26941. doi.org/10.1371/journal.pone.0026941
- Beaumont AR, Fairbrother JE, Hoare K, 1995, Heredity, 75(3):256–266, doi:10.1038/hdv.1995.133.

